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1	828337	DNA near5 isolat\$ or extract\$ or purify\$	USPAT; US-PGPUB; DERWENT	2003/07/02 11:36
2	4246	(DNA near5 isolat\$ or extract\$ or purify\$) and high adj1 salt	USPAT; US-PGPUB; DERWENT	2003/07/02 11:37
3	1354	((DNA near5 isolat\$ or extract\$ or purify\$) and high adj1 salt) and lysis	USPAT; US-PGPUB; DERWENT	2003/07/02 11:38
4	869	((((DNA near5 isolat\$ or extract\$ or purify\$) and high adj1 salt) and lysis) and reagent	USPAT; US-PGPUB; DERWENT	2003/07/02 11:39
5	797	((((DNA near5 isolat\$ or extract\$ or purify\$) and high adj1 salt) and lysis) and reagent) and biological	USPAT; US-PGPUB; DERWENT	2003/07/02 11:40

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L4 ANSWER 1 OF 2 MEDLINE
AN 1999247832 MEDLINE
DN 99247832 PubMed ID: 10232847
TI Osmotically driven radial diffusion of single-stranded DNA fragments on an agarose bed as a convenient measure of DNA strand scission.
AU Sestili P; Cantoni O
CS Istituto di Farmacologia e Farmacognosia and Centro di Farmacologia Oncologica Sperimentale, Universita di Urbino, Italy..
sestili@fis.uniurb.it
SO FREE RADICAL BIOLOGY AND MEDICINE, (1999 Apr) 26 (7-8) 1019-26.
Journal code: 8709159. ISSN: 0891-5849.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 199906
ED Entered STN: 19990712
Last Updated on STN: 19990712
Entered Medline: 19990623

L4 ANSWER 2 OF 2 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC.DUPLICATE 1
AN 1984:175360 BIOSIS
DN BA77:8344
TI FLUOROMETRIC QUANTIFICATION OF DNA IN CELLS AND TISSUE.
AU DOWNS T R; WILFINGER W W
CS DEP. PHYSIOL., UNIV. CINCINNATI, COLL. MED., 213 BETHESDA AVE.,
CINCINNATI, OHIO 45267.
SO ANAL BIOCHEM, (1983) 131 (2), 538-547.
CODEN: ANBCA2. ISSN: 0003-2697.
FS BA; OLD
LA English

=> d 14 1-2 abs

L4 ANSWER 1 OF 2 MEDLINE
AB The present study describes the development and characterization of a novel technique, the alkaline-halo assay, for the assessment of DNA single strand breakage in mammalian cells. This technique allows the measurement of DNA lesions at the single cell level and presents the additional advantages of being rapid, sensitive, virtually costless and environmentally friendly, because it does not require the use of isotopes. The alkaline halo assay involves a series of sequential steps in which the cells are first treated, then embedded in melted agarose and spread onto microscope slides that are incubated for 2 min at ice-bath temperature to allow complete geling. The slides are then incubated for 20 min in a **high salt alkaline lysis solution**, for an additional 15 min in a hypotonic alkaline solution and, finally, for 10 min in ethidium bromide. Under these conditions, single-stranded DNA fragments spread radially from the nuclear cage and generate a fluorescent image that resembles a halo concentric to the nucleus remnants. The area of the halos increased at increasing levels of DNA fragmentation and this process was associated with a progressive reduction of areas of the nuclear remnants. These events were conveniently monitored with a fluorescence microscope and quantified by image processing analysis. The sensitivity of the alkaline-halo assay, which is based on the osmotically driven radial diffusion of single-stranded DNA fragments through agarose pores, is remarkably similar to that of the widely used alkaline elution and comet assays.

L4 ANSWER 2 OF 2 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC.DUPLICATE 1
AB The validation of a simple and rapid DNA solubilization procedure is described. Quantitative **extraction** of intact, polymerized DNA was achieved by [rat] cell lysis or tissue homogenization in an

ammonium hydroxide-Triton X-100 solution. The solubilization procedure inactivates endogenous DNase and increases the fluorescence-enhancement activity of the **extracted** DNA, thereby eliminating the need for enzyme treatment or exposure to **high salt solutions**. The **extracts** can be utilized directly in a sensitive fluorescence-enhancement assay with bisbenzimidazole (Hoechst 33258) reagent. Estimates of DNA cell content were unaffected by the number of cells lysed or the volume of lysate employed in the assay. In all cases, the solubilized DNA estimates were linear and parallel to the bovine DNA standard. The optimum range for estimation of DNA in this assay is 5-150 ng. In addition, estimates of DNA obtained with this method and the standard diphenylamine assay were in excellent agreement. This simple, 1-step DNA **extraction** procedure can be utilized in conjunction with Hoechst reagent to obtain quantitative estimates of DNA levels in cell or tissue **extracts**.

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FULL ESTIMATED COST		21.33	21.54

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E7	1 HEATH ELIZABETH I/AU
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E15	1 HEATH FRACICA L A/AU
E16	19 HEATH G/AU
E17	9 HEATH G A/AU
E18	18 HEATH G B/AU
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E20	9 HEATH G E/AU
E21	5 HEATH G F/AU
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L5	1 ("HEATH ELLEN M"/AU)

=> DIS L5 1 IBIB ABS

L5 ANSWER 1 OF 1 MEDLINE
ACCESSION NUMBER: 2002125907 MEDLINE
DOCUMENT NUMBER: 21849689 PubMed ID: 11860298
TITLE: Utilizing genomic DNA purified from clotted blood samples for single nucleotide polymorphism genotyping.
AUTHOR: Adkins Karissa K; Strom Daniel A; Jacobson Thomas E; Seemann Cara R; O'Brien Darin P; Heath Ellen M
CORPORATE SOURCE: Gentra Systems Inc, Minneapolis, MN 55441, USA.
SOURCE: ARCHIVES OF PATHOLOGY AND LABORATORY MEDICINE, (2002 Mar) 126 (3) 266-70.
Journal code: 7607091. ISSN: 0003-9985.
PUB. COUNTRY: United States
DOCUMENT TYPE: (EVALUATION STUDIES)
Journal; Article; (JOURNAL ARTICLE)
LANGUAGE: English
FILE SEGMENT: Abridged Index Medicus Journals; Priority Journals
ENTRY MONTH: 200203
ENTRY DATE: Entered STN: 20020226
Last Updated on STN: 20020403
Entered Medline: 20020327

AB CONTEXT: Linking single nucleotide polymorphisms to disease etiology is expected to result in a substantial increase in the number of genetic tests available and performed at clinical laboratories. Whole blood serves as the most common DNA source for these tests. Because the number of blood samples rises with the number of genetic tests performed, alternative DNA sources will become important. One such alternative source is clotted blood, a by-product of serum extraction. Efficiently using an already procured blood sample would limit the overall number of samples processed by clinical laboratories. OBJECTIVE: To determine if DNA purified from clotted blood can be effectively used for single nucleotide polymorphism genotyping. DESIGN: DNA was purified from the clotted blood of 15 donors. Single nucleotide polymorphism genotyping for the methylenetetrahydrofolate reductase and factor V Leiden mutations was performed with each DNA sample by 2 independent methods. RESULTS: High-quality DNA was obtained from each of the 15 individual clotted blood samples as demonstrated by UV spectrophotometric analysis, gel electrophoresis, and polymerase chain reaction amplification. The DNA was used successfully to obtain genotype data from both the methylenetetrahydrofolate reductase and factor V single nucleotide polymorphism assays for all samples tested. CONCLUSIONS: Clotted blood is a clinically abundant sample type that can be used as a source of high-quality DNA for single nucleotide polymorphism genotyping.

=> d his

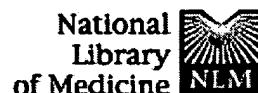
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L1 3627036 S DNA (2A) ISOLAT? OR EXTRACT? OR PURIF?
L2 347 S L1 AND HIGH (1A) SALT (2A) SOLUTION
L3 4 S L2 AND LYSIS
L4 2 DUP REM L3 (2 DUPLICATES REMOVED)

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L5 1 S (E8)



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